# Applications and issues of large scale transcriptome profiling experiments

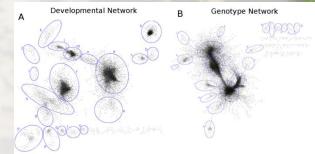




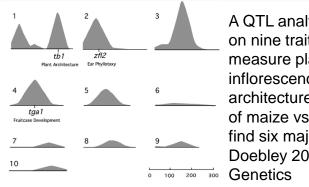
# Outline

- Co-expression and expression conservation
  - Reshaping of the maize transcriptome by domestication (Swanson-Wagner et al PNAS 2012)
  - Variation among networks
  - RNAseq vs microarray
- Enabling usage of co-expression networks to study natural variation
  - eQTL hotspots
  - Phenotypic QTL
- Integration of transcriptome and epigenome





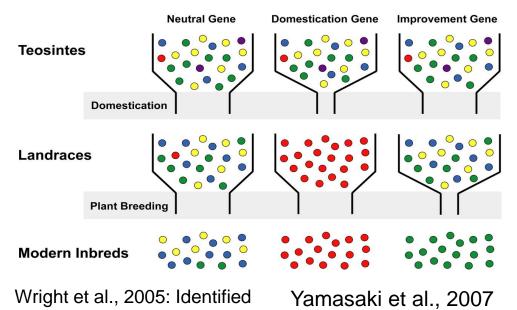
### Identification of loci involved in domestication

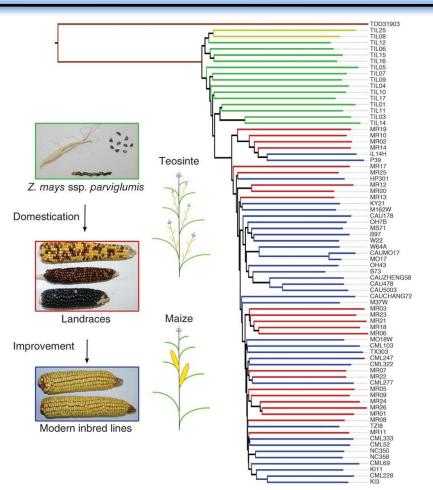


~30 targets of domestication

A QTL analysis focused on nine traits that measure plant and inflorescence architecture in a cross of maize vs teosinte find six major effect loci: Doebley 2004 Ann Rev Genetics

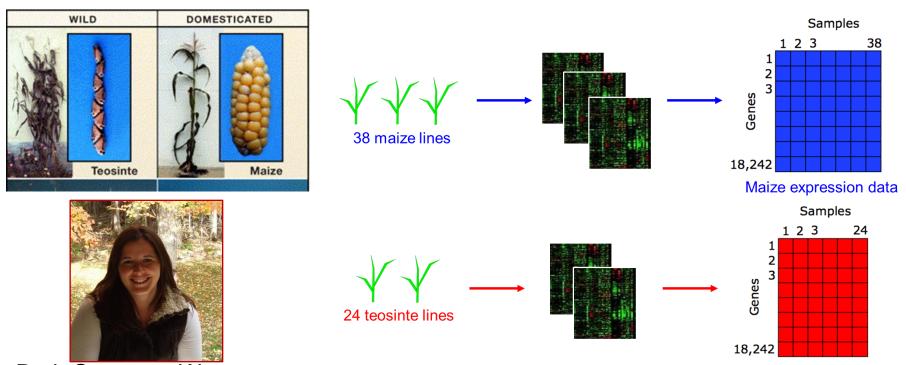
#### Genomic scans for selection (diversity scans)





Hufford et al., 2012 Re-sequence 75 genomes Identified ~500 selected regions (1754 genes)

#### **Collection of expression data**

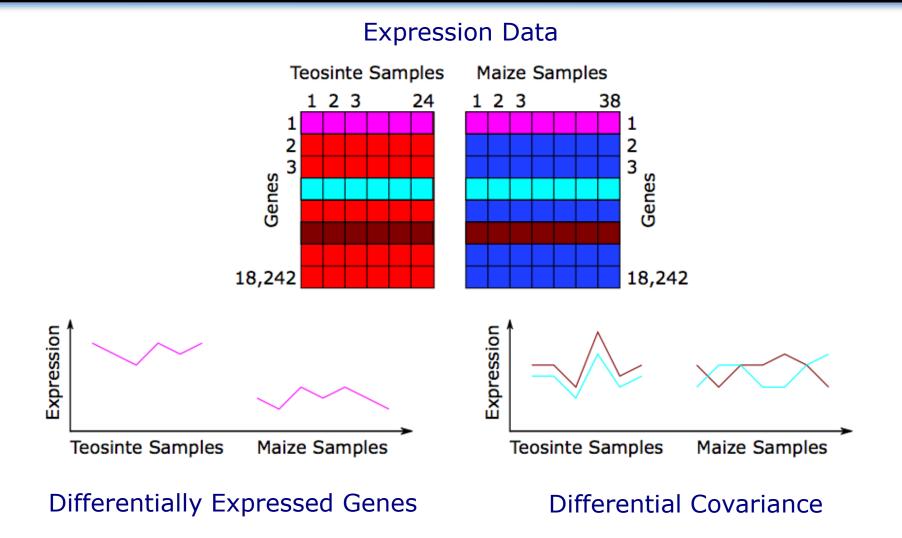


Ruth Swanson-Wagner

Teosinte expression data

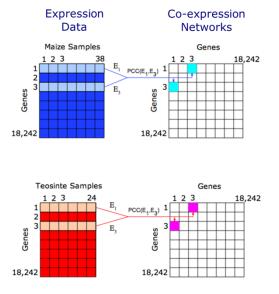
- Maize: 38 genotypes assayed (23 NAMs and other diverse maize inbreds)
- Teosinte: 24 genotypes profiled (7 TILs and 17 "wild" individuals)
- Seedling expression assayed by using custom NimbleGen array with 3-4 probes each for ~32,500 4a.53 filtered gene set

### Finding differences in expression data

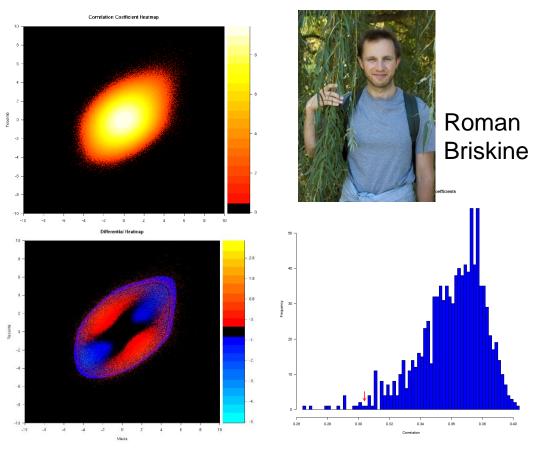


#### **Re-wiring of transcriptome in maize**

- Generate co-expression networks in maize and teosinte
- Assess network similarity and per-gene expression conservation (EC)

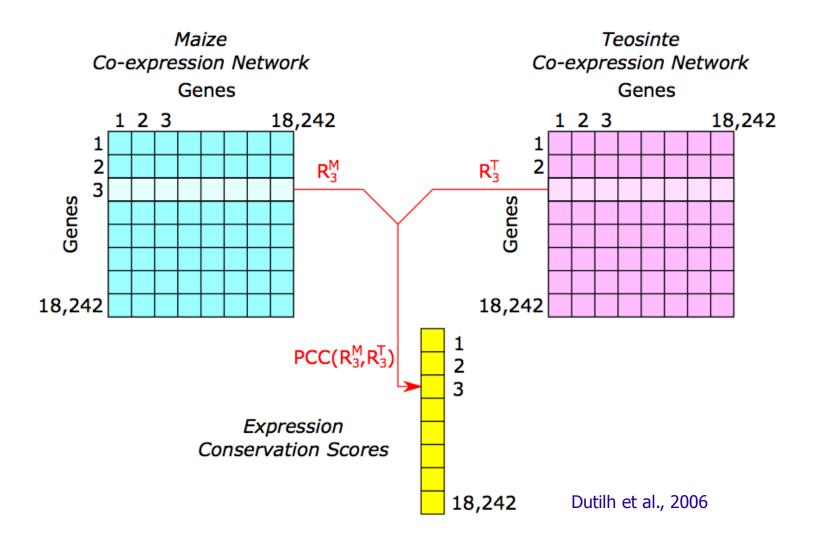


- Co-expression network records similarity between each pair of gene expression profiles.
- Fisher transformation and normalization (Huttenhower et al., 2006)

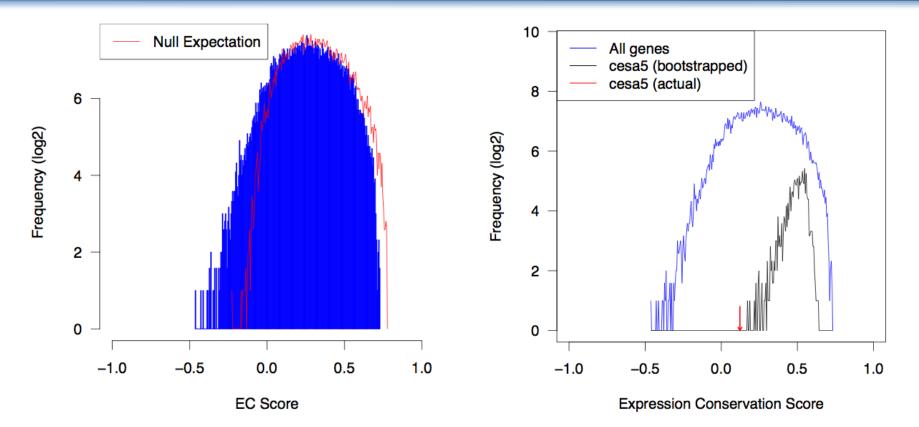


Transcriptome is significantly re-wired

# Expression conservation score measures similarity between gene's co-expression profiles in two networks



# Identification of genes with significant differences in EC

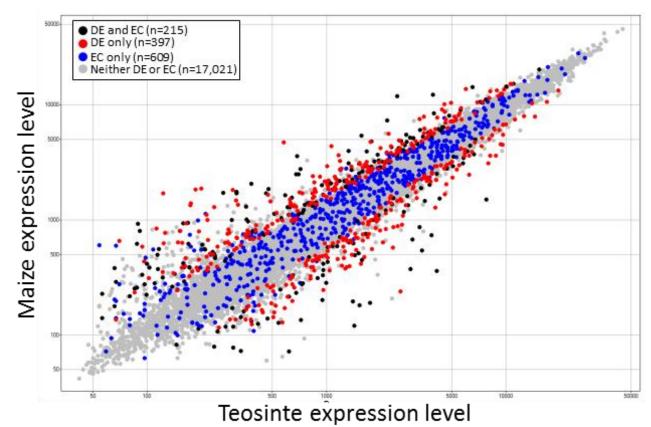


$$z = \frac{EC - \mu_{null}}{\sigma_{null}}$$

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#### EC and DE approaches identify different expression changes

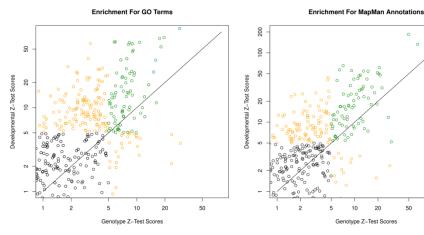
- 18,224 expressed genes assessed
- 612 DE genes (enriched for targets of selection)
- 824 AEC genes
- 215 in common (enriched for targets of selection)

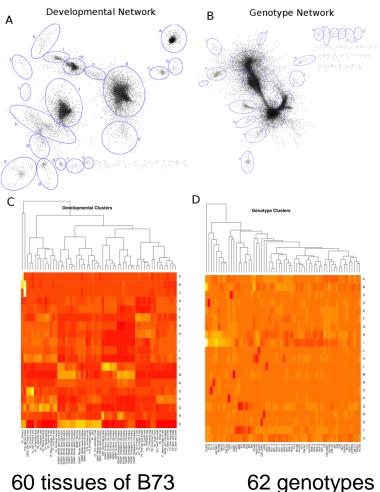


### **Co-expression networks**

- **Co-expression analysis** identifies genes with similar patterns of expression: **Relies upon variation in** gene expression
- Should we be using the "kitchen sink" approach or developing multiple networks?

Genotype Z-Test Scores





Swanson-Wagner et al 2012

electron transport chain

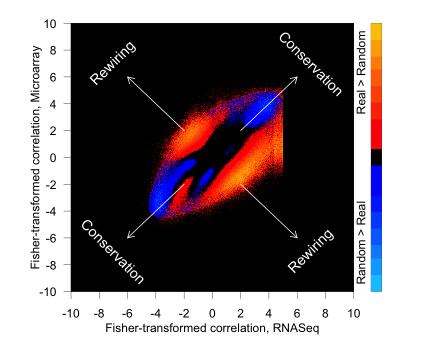
glucose metabolism

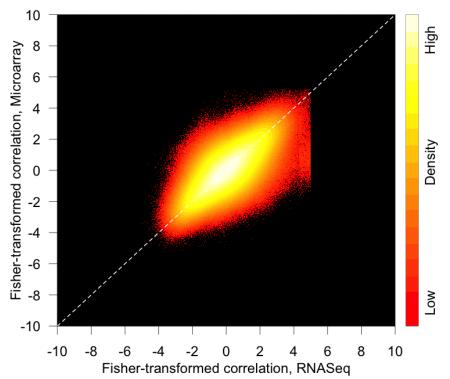
response to biotic stimulus

60 tissues of B73 Sekhon et al., 2011 protein catabolic processes organic substance transport cell wall modification

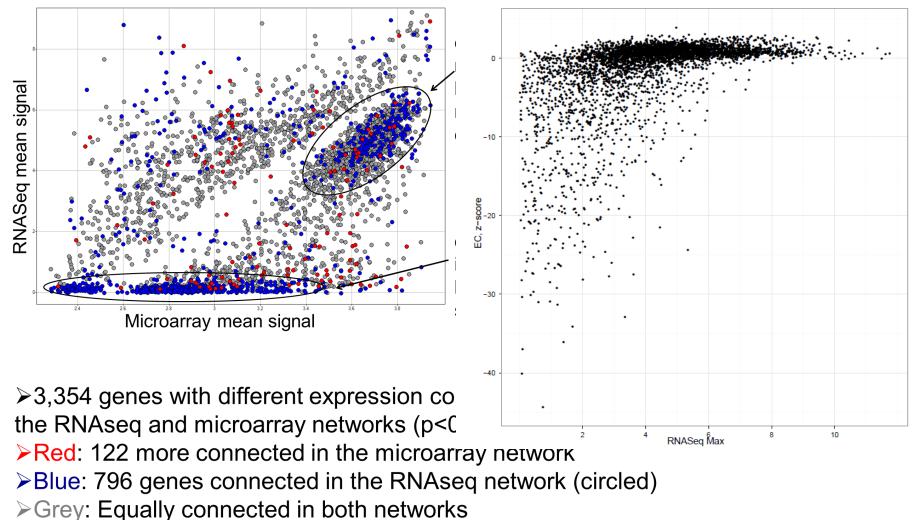
### **Co-expression networks: RNAseq vs microarray**

- Comparison of microarray and RNAseq data: Co-expression
  - 18 samples from different tissues of B73
  - Selected 19,328 "expressed" genes from microarrays





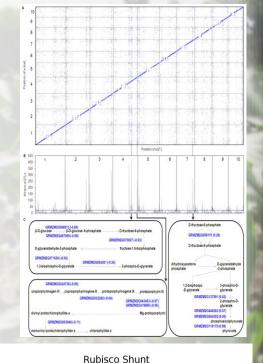
### **Expression conservation: RNAseq vs microarray**

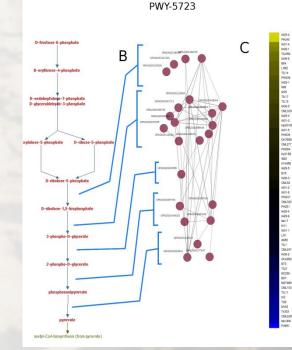


Maximum expression level vs. Expression Conservation z-score

# Outline

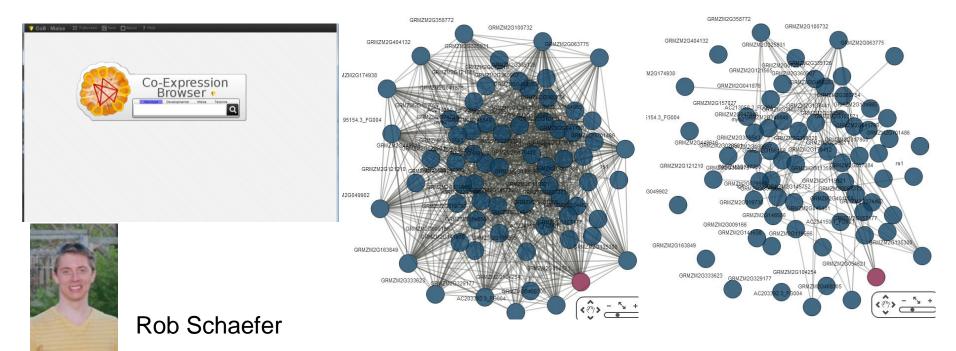
- Co-expression and expression conservation
- Enabling usage of co-expression networks to study natural variation
  - Simple user queries of networks
  - eQTL hotspots
  - Phenotypic QTL
- Integration of transcriptome and epigenome



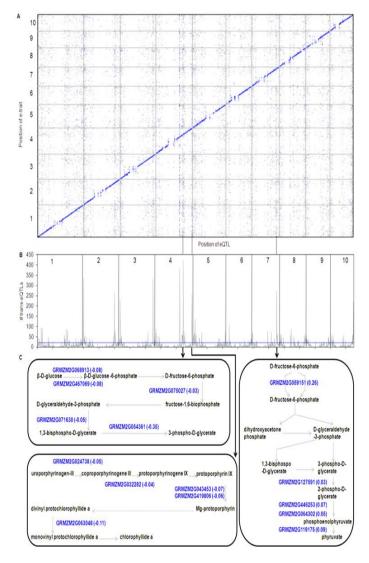


# COB: A viewer to query co-expression networks with genes and coordinates

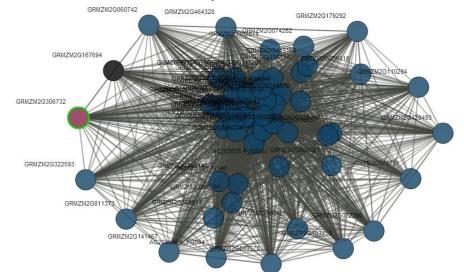
- http://csbio.cs.umn.edu/cob/
- Allows user to query various networks with gene(s) and then to visualize genomic coordinates or overlap between networks



### **Co-expression networks: Trans-eQTL hotspots**



- Trans-eQTL "hotspots" identified using RNAseq analysis of ~100 RILs
- Determine whether "targets" are coexpressed in other genotypes or tissues
- Ask whether genes within hotspot are in same network
- Several examples in which putative TF within hotspot shows co-expression with network in other samples

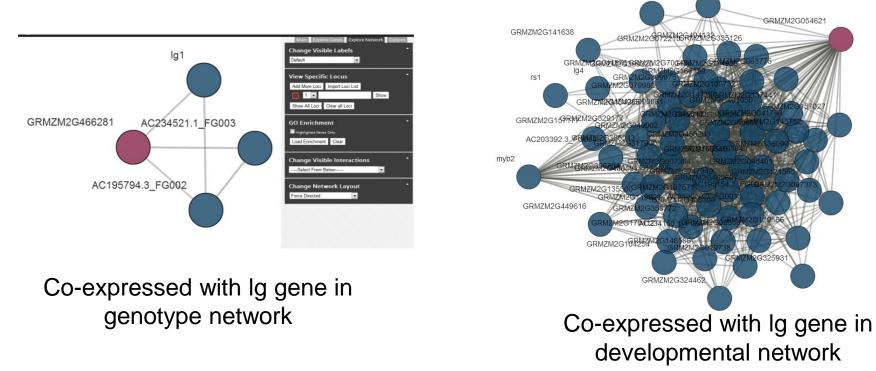


Gene from trans-eQTL hotspot co-expressed with many targets

### **Co-expression networks: Phenotypic QTL**

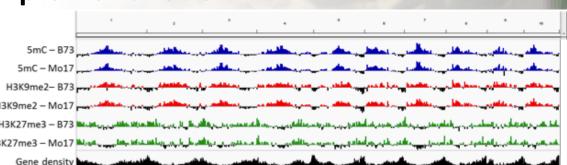
- Tian et al (2011) identified ~30 QTL for leaf angle by joint linkage analysis
  - Also performed GWAS
- Two classical maize mutants; Ig1 and Ig2 likely are molecular bases for two of the QTL (and have significant SNP associations)
- Rest are unknown
- Query co-expression networks to identify genes co-expressed with Ig genes and located within QTL

GRMZM2G009166



# Outline

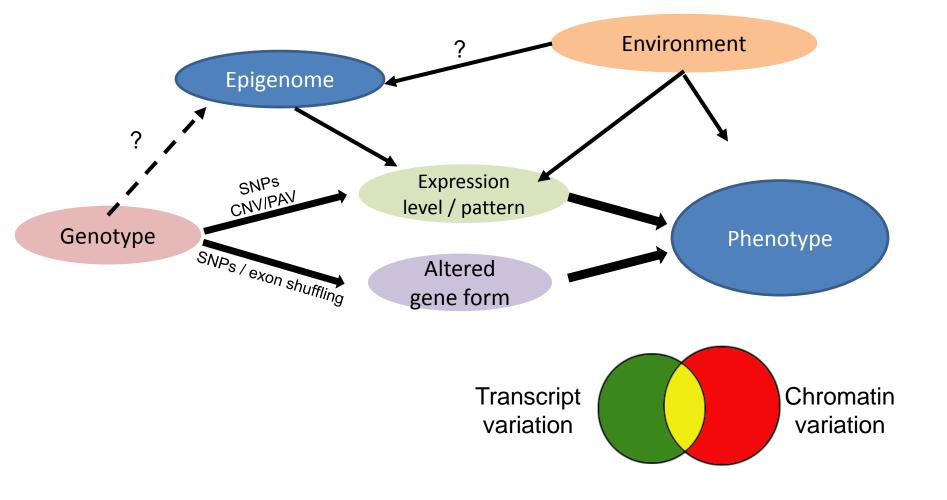
- Co-expression and expression conservation
- Enabling usage of co-expression networks to study natural variation
- Integration of transcriptome and epigenome
  - Different data types
  - How to isolate contribution of epigenome to transcriptome variation



Me

# Transcriptome profiling provides critical information for understanding phenotype

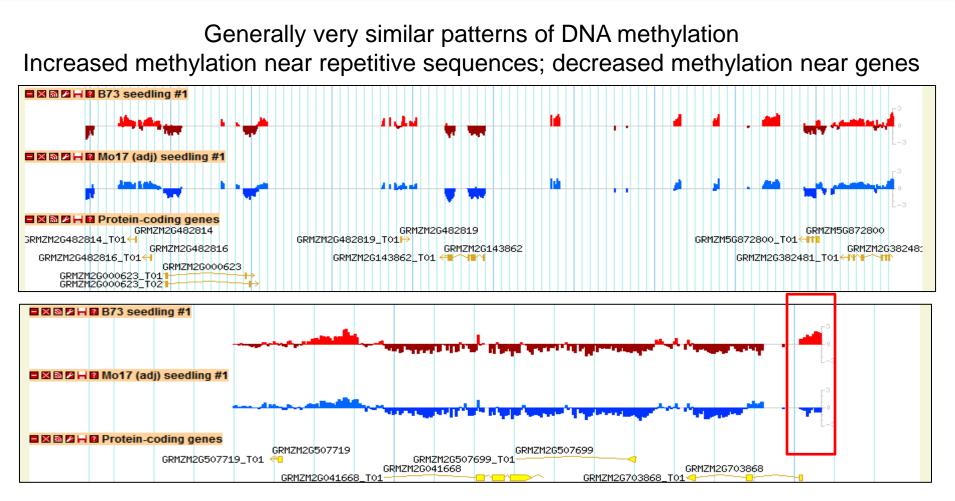
• What proportion of expression level variation is attributable to epigenome?



### **Data and questions**

- Data types:
  - meDIP-chip (DNA methylation) [n=~140 profiles]
  - ChIP-chip (H3K9me2; H3K27me3) [n=~75 profiles]
  - RNAseq 120 samples (20-25 million reads each)
- Samples
  - Five tissues for two genotypes
  - 1 tissue for 25 genotypes
- Identification of initial variation (two samples with replicates) easy
- How to collapse and classify variation in large population more difficult
- Overlap? (lots of samples, not requiring complete correlation)
  - Chromatin marks and expression
  - Chromatin marks and SNPs

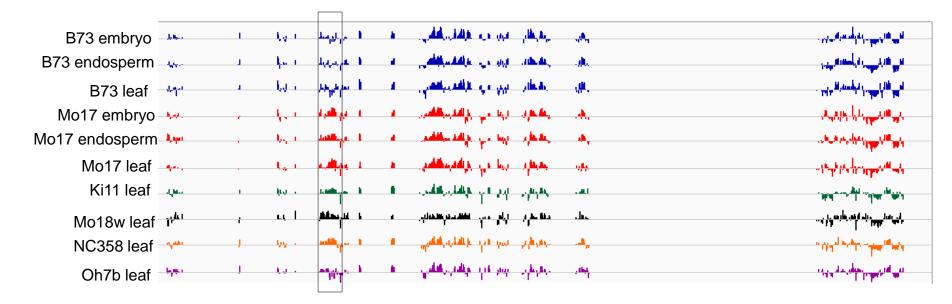
### Limited variation for DNA methylation patterns



Regions with extremely different methylation profiles can also be found ~1000 DMRs in B73 vs Mo17

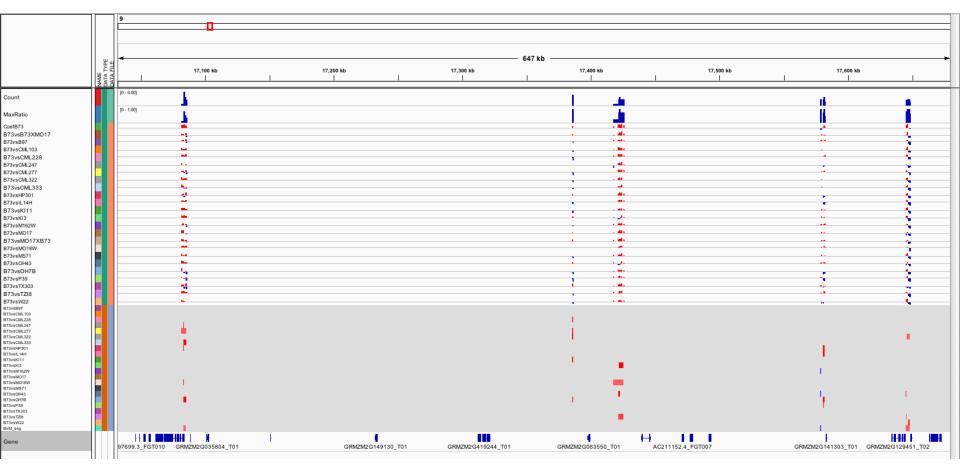
### What happens in other maize genotypes?

- DNA methylation patterns are generally quite similar among genotypes and tissues.
- However, there are ~1000 DMRs between any two genotypes.
- Variation frequently acts equally upon all tissues.



### What happens in other maize genotypes?

- Call DMRs between two genotypes
- Need tools for simultaneously defining regions and classifying among all genotypes



### **Once DMRs are found: Causes and Effects**

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 What is causing chromatin change? Is it associated with SNPs? Rare phenotype problem

 Does the chromatin change cause an expression change? What about partial correlations?

## Summary

- Making sense of differences among populations
  - Co-expression and expression conservation
- Enabling usage of –omics datasets (transcriptome, epigenome, etc)
  - Interrogation tools
  - Visualization tools
- When is enough enough?
  - Allele-specific expression analysis

# Thanks!

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